

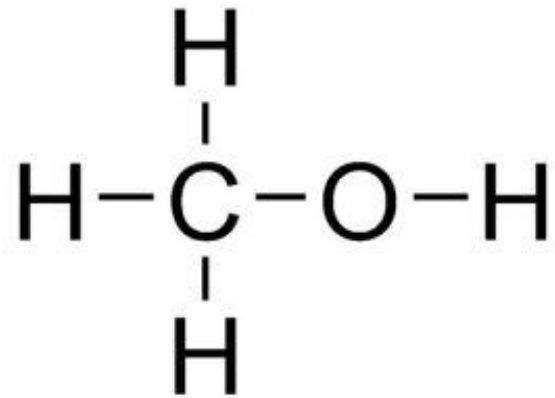
Journal Club

2/6/2020

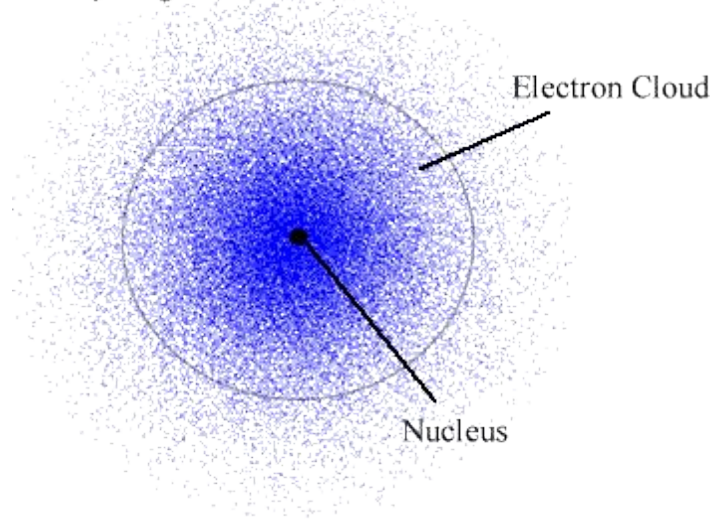
Maria Perica

Outline

- MRS 101
- Spectral quality
- LCModel output
- Proposed updates to procedure
- Metabolites we collect



Hydrogen Atom Electron Cloud Model



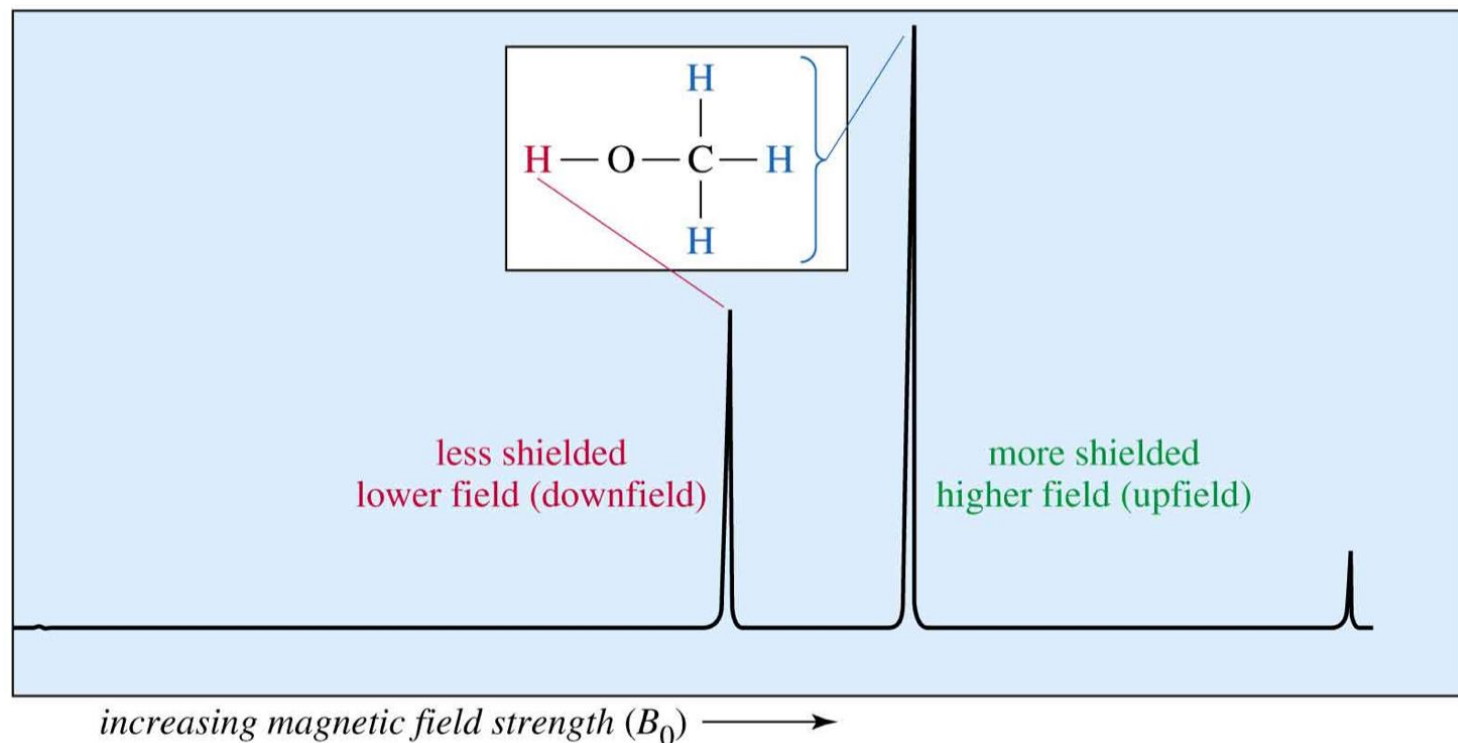
MRS aka Chemical Shift Imaging

- Measure metabolites noninvasively in vivo
 - Organic compound structure determination
 - Same principles as conventional MRI
- Magnetic field that a molecule experiences depends on chemical structure and the environment in which it is
- Slightly different effective field
- CS gives rise to the spectrum

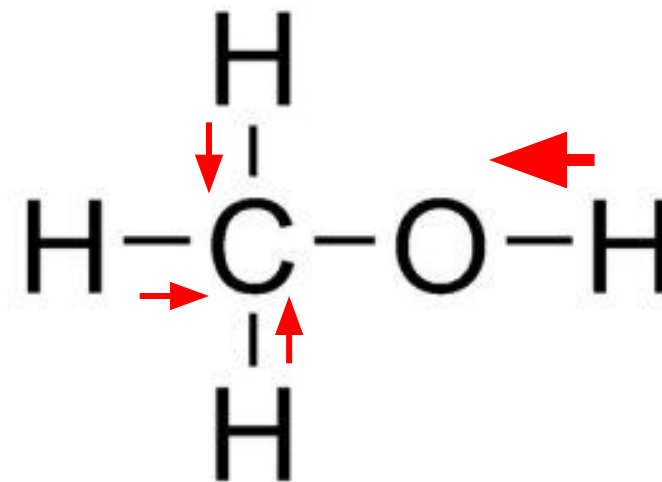
Shielding and **deshielding**: nucleus "experiences" weaker or stronger magnetic field as a result of the electron density

Upfield and **downfield**

- More shielded \square stronger field must be applied

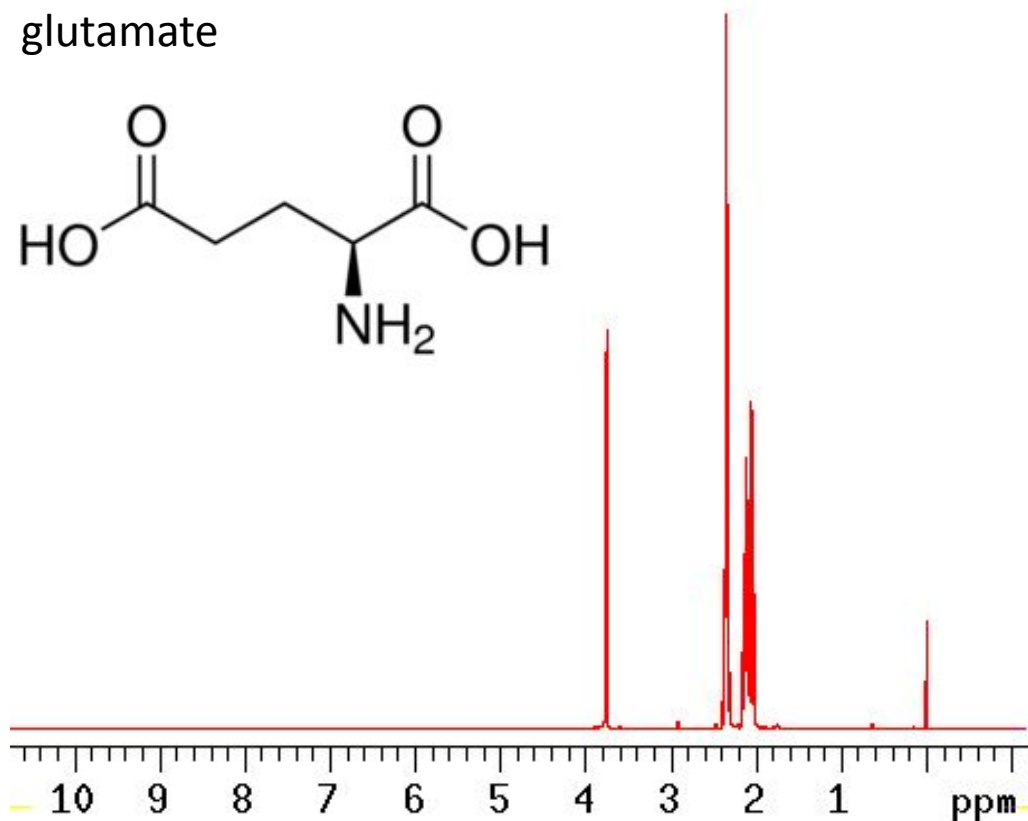


Electronegativity also influences shielding



From spectrum to molecule

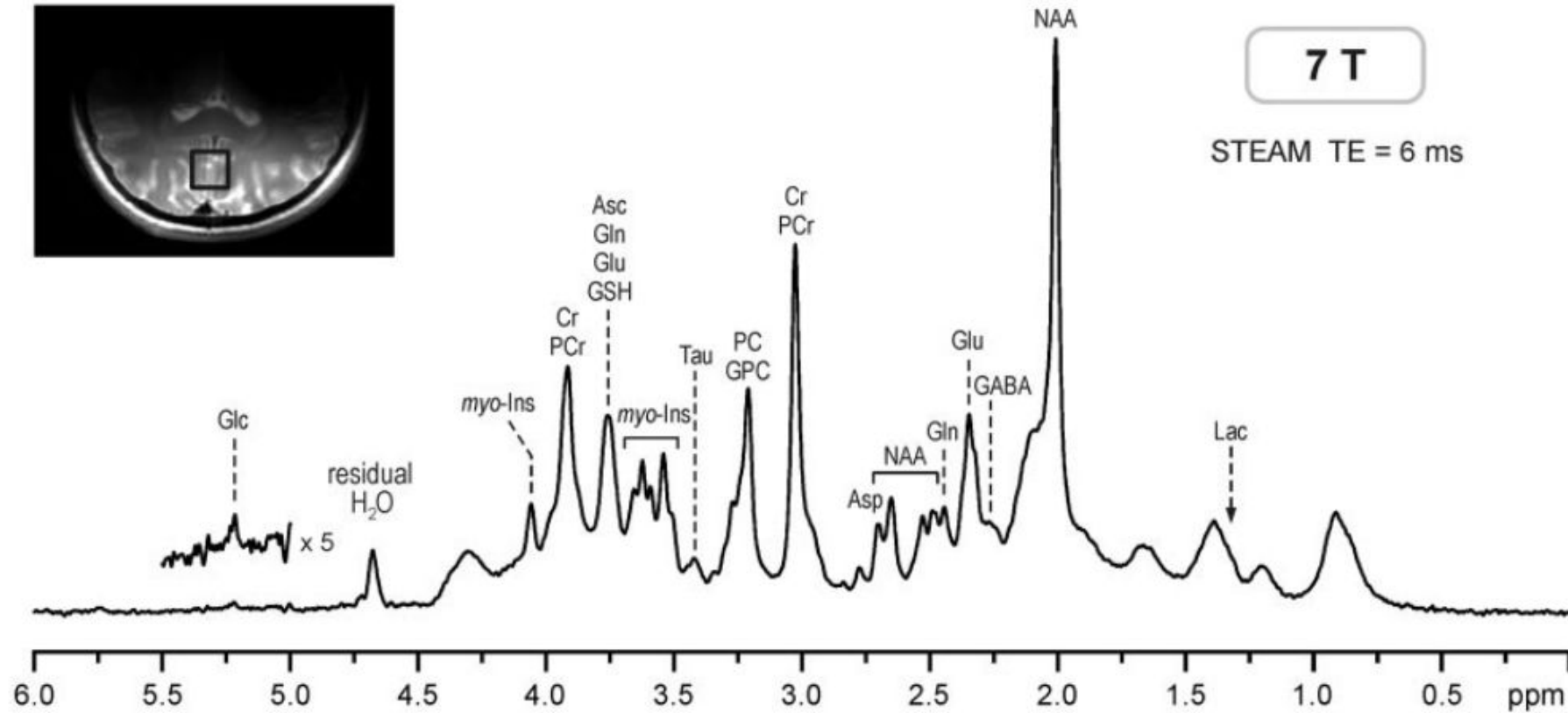
- Location of peaks shielding
- Number of peaks Number of H
- Intensity of signal Number of that type of H
- Signal splitting Number of protons on adjacent atoms
- Integral Concentration

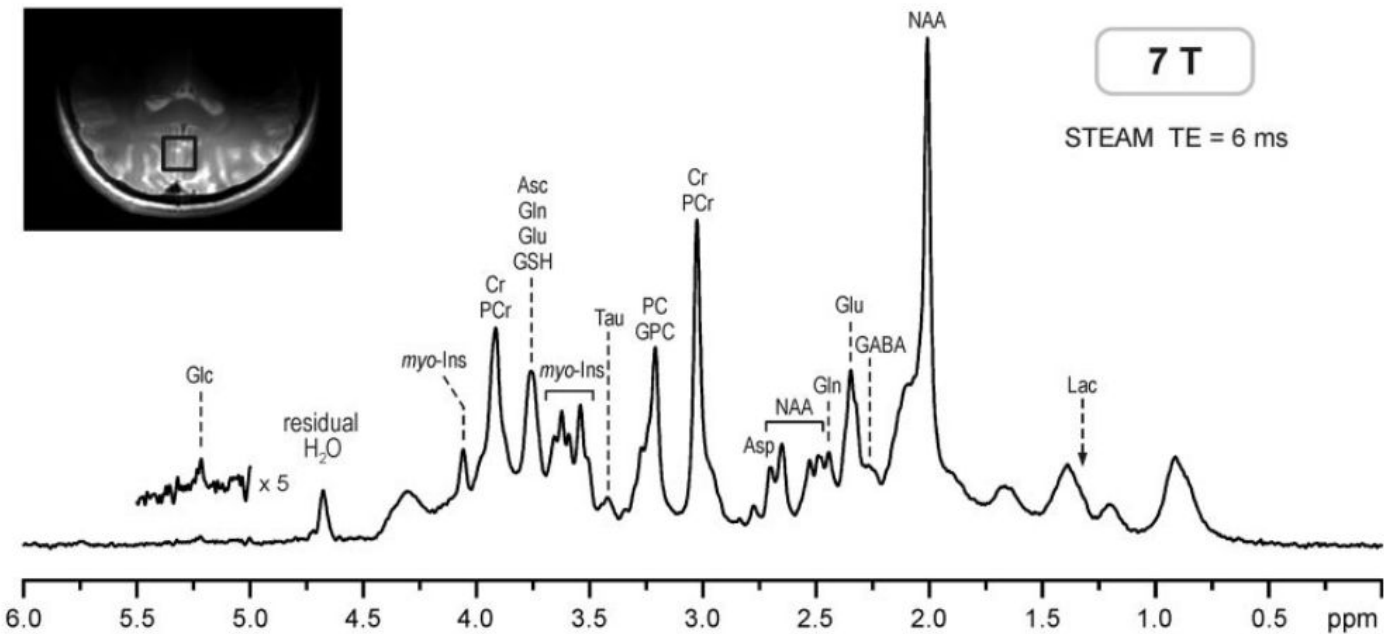
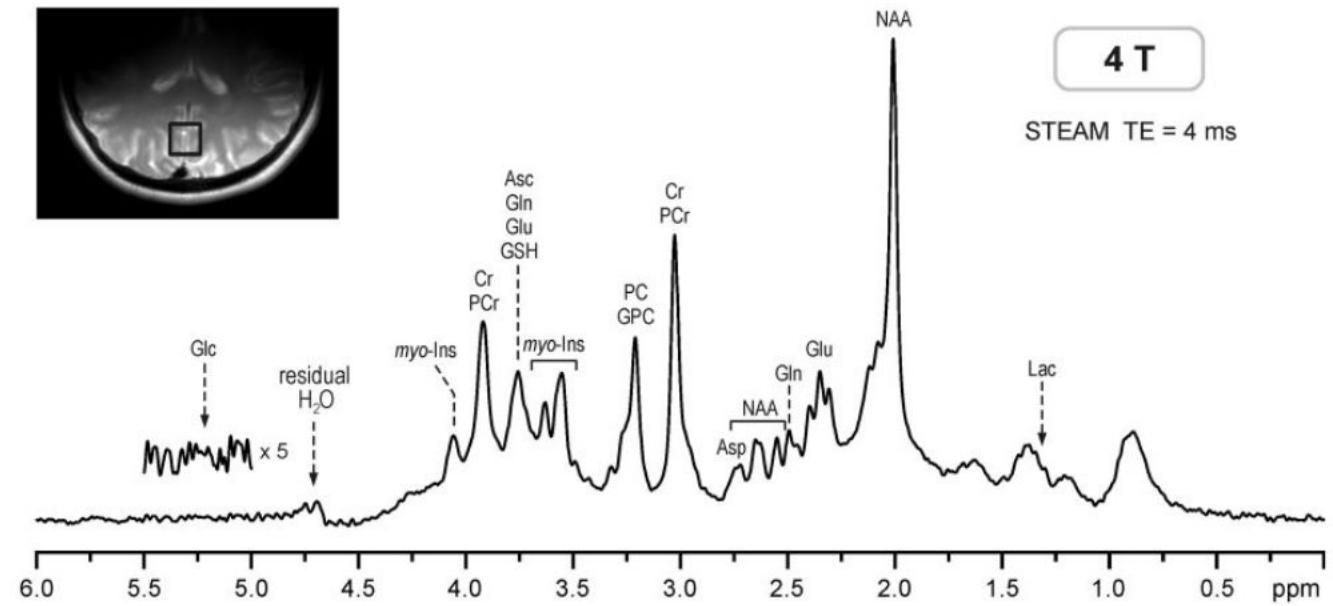


A dark blue, irregularly shaped graphic with a splatter effect, containing white text. The graphic is centered on a white background and has a rough, hand-painted appearance with some lighter blue and white speckles around its edges.

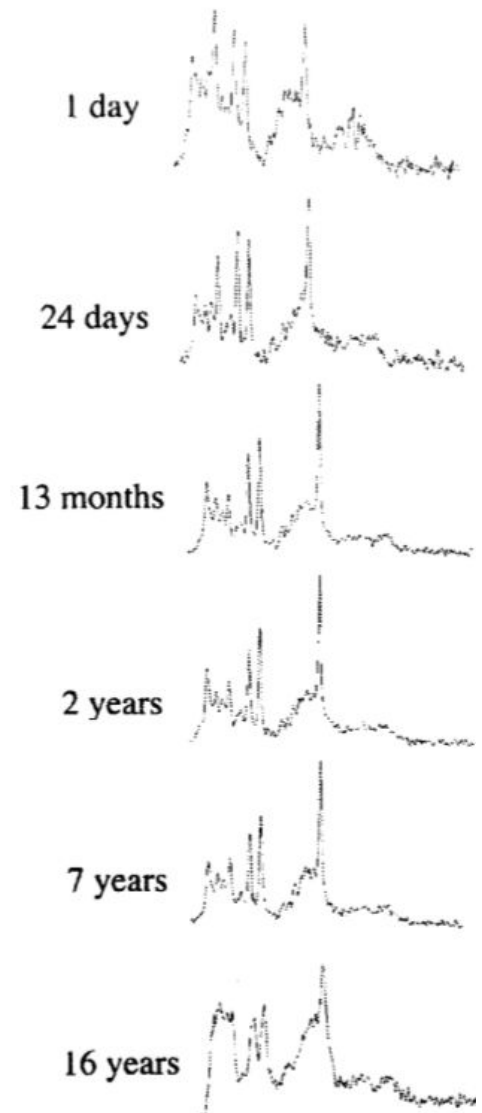
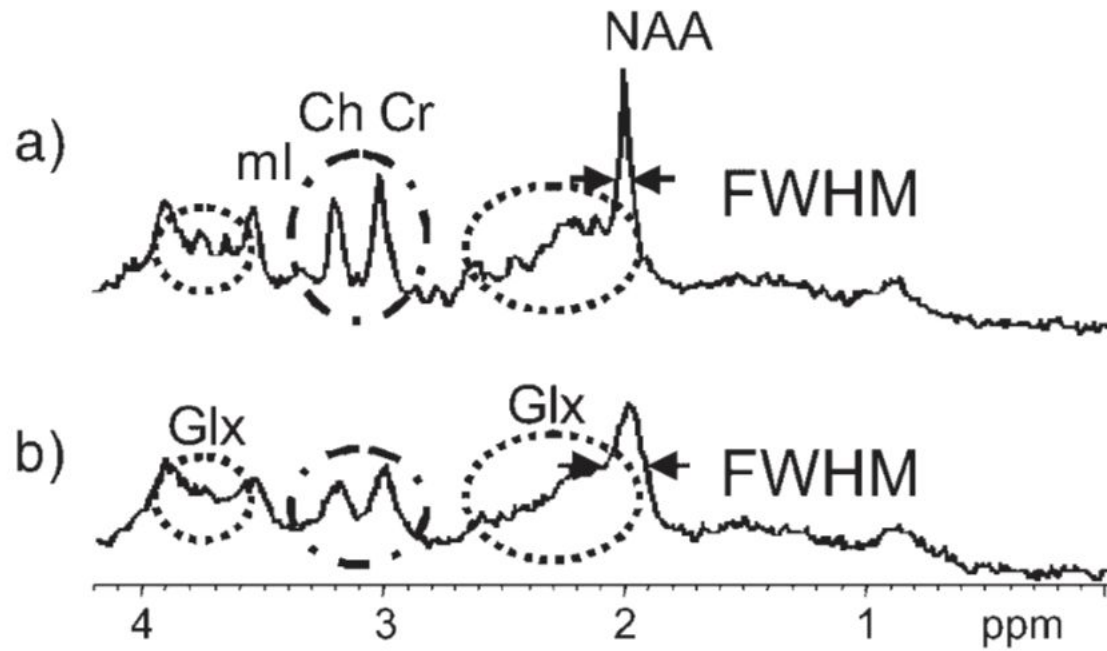
Gallery of spectral quality and artifacts

Good spectrum

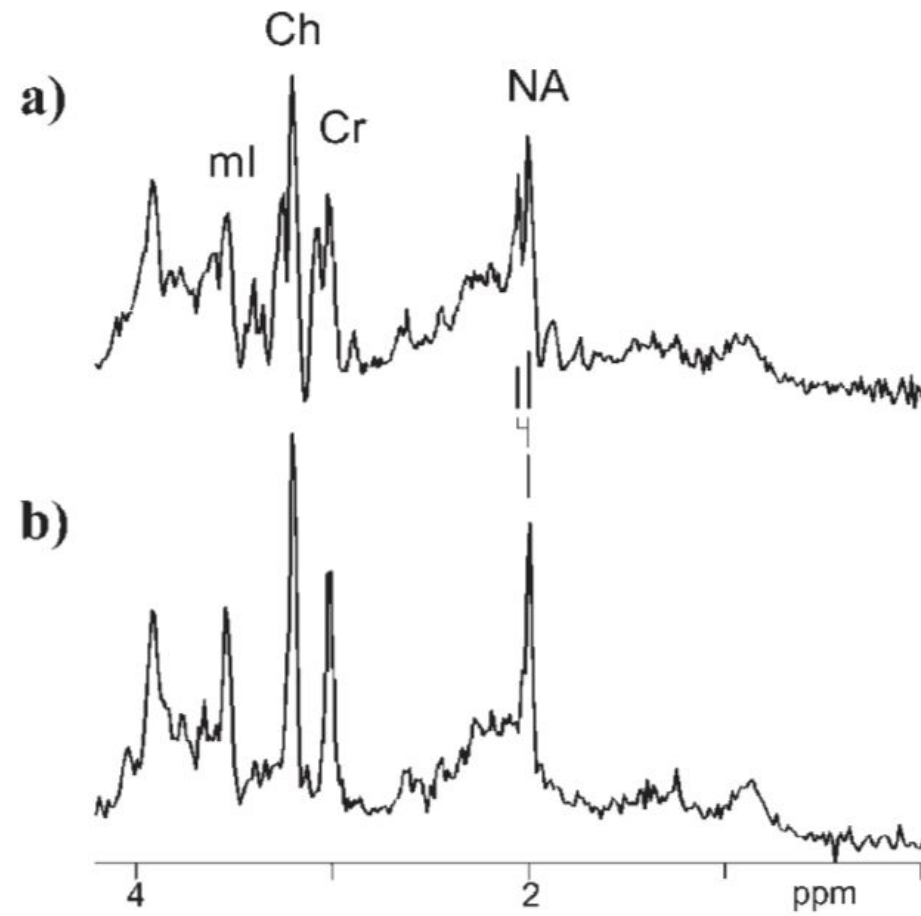




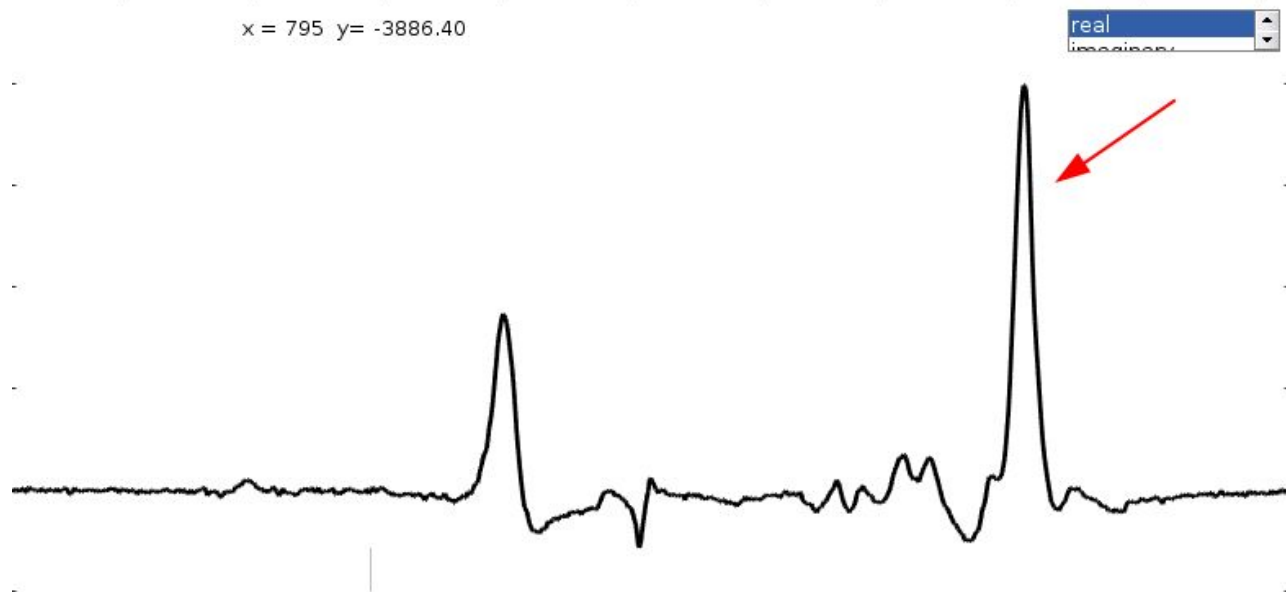
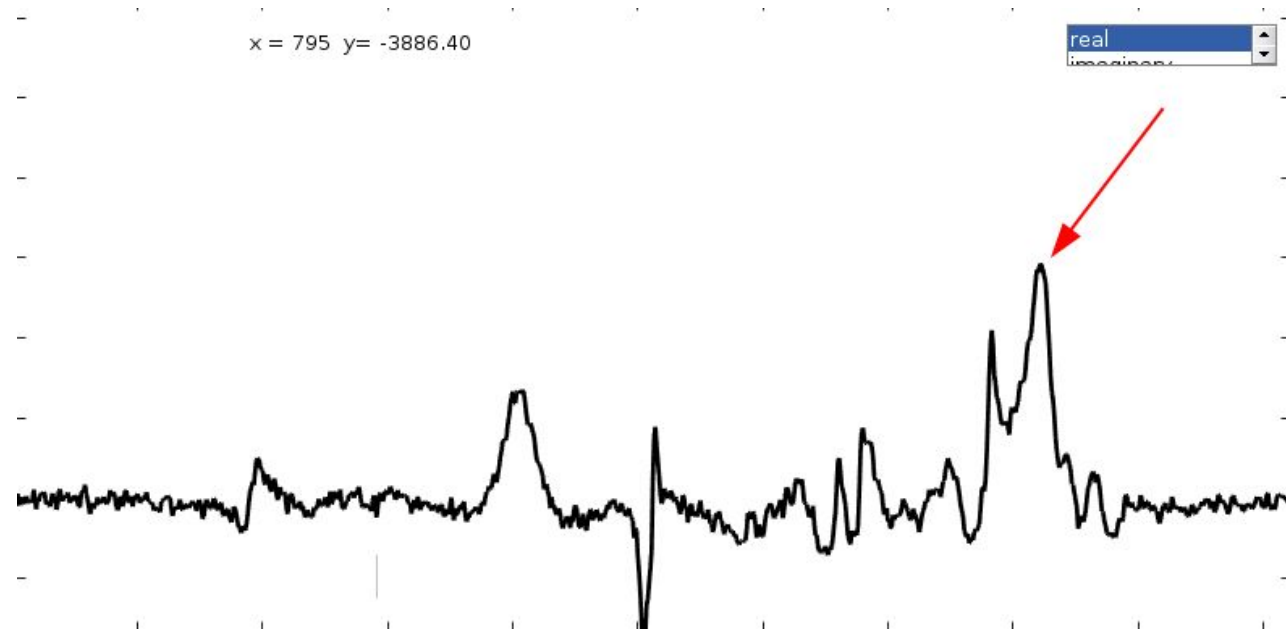
Linewidth



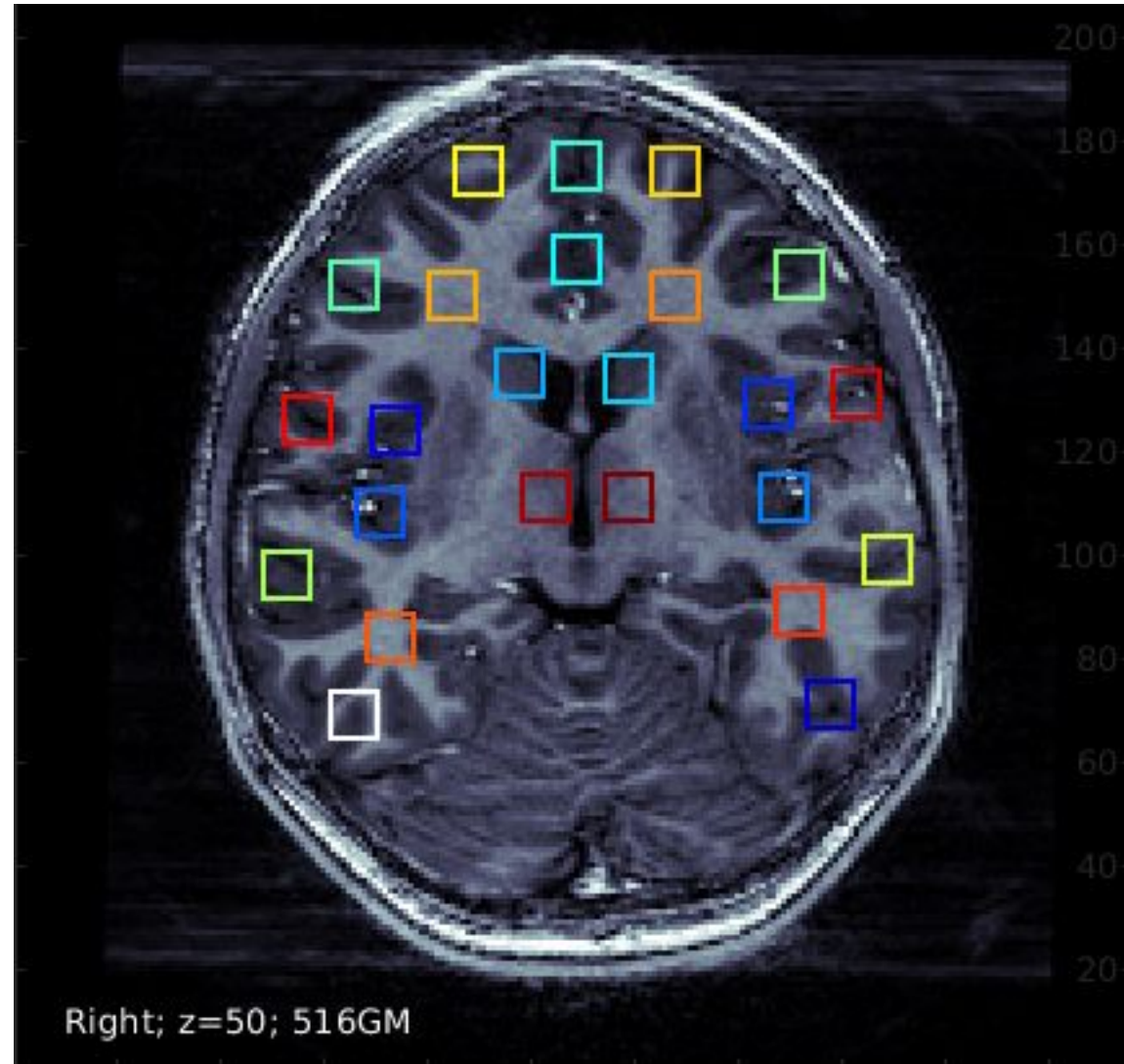
Motion



Lipids

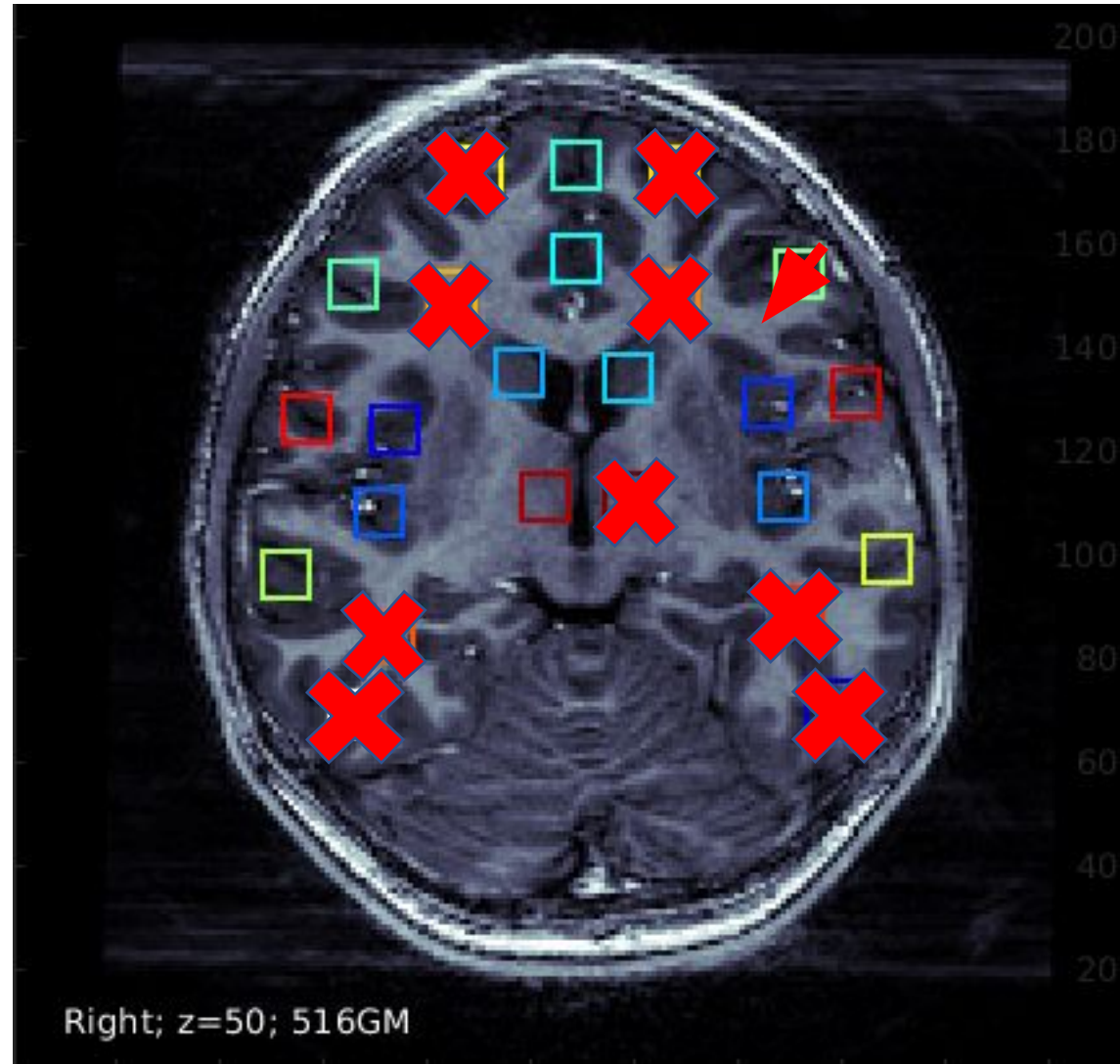


Coordinates: Before



Proposed updates

2. Remove SFG, WM voxels, one thalamus voxel because they are too close together and not statistically independent



1. Move all peripheral voxels away from outer-volume lipids

3. Remove MOG voxels because bad data quality as a result of not optimized acquisition

LCModel

/home/MRSI/data/Luna/20190507/11664_20180712-20180712Luna1/spectrum.113.150

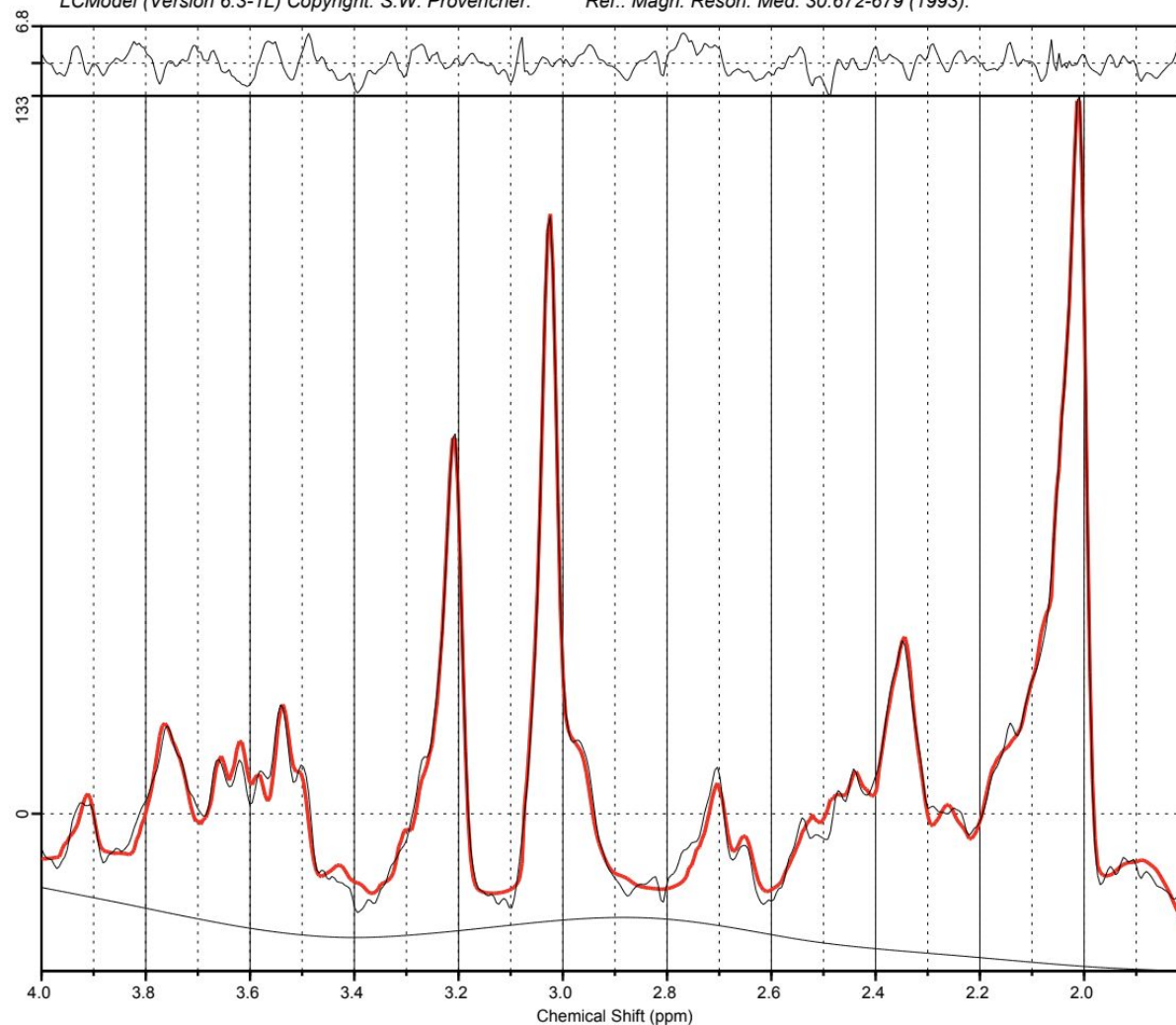
Data of: Department of Radiology, University of Pittsburgh

LCModel (Version 6.3-1L) Copyright: S.W. Provencher.

Ref.: Magn. Reson. Med. 30:672-679 (1993).

09-May-2019 14:22

1. residuals



Conc.	%SD	/Cre	Metabolite
0.000	999%	0.000	Asp
13.916	25%	0.144	Cho
96.725	2%	1.000	Cre
40.217	6%	0.416	GABA
25.042	21%	0.259	Glc
48.439	6%	0.501	Gln
134.241	3%	1.388	Glu
14.883	23%	0.154	GPC
27.931	7%	0.289	GSH
125.851	3%	1.301	mI
114.215	3%	1.181	NAA
20.716	16%	0.214	NAAG
15.923	17%	0.165	Tau
18.453	11%	0.191	-CrCH2
28.800	2%	0.298	GPC+Cho
134.931	3%	1.395	NAA+NAAG
182.679	3%	1.889	Glu+Gln

91.066 15% 0.941 MM20

DIAGNOSTICS
1 warning MYBASI 41

MISCELLANEOUS OUTPUT
FWHM = 0.035 ppm S/N = 30
Data shift = -0.035 ppm
Ph: 17 deg 19.0 deg/ppm

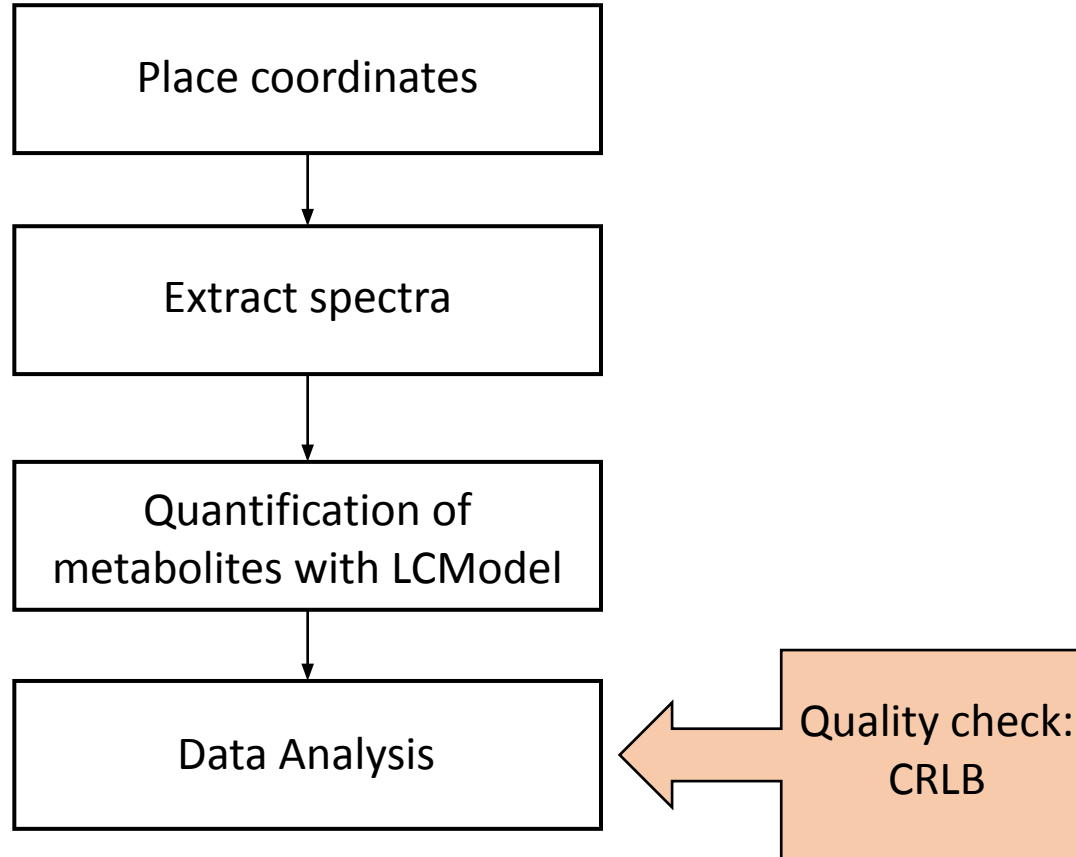
INPUT CHANGES
LCSV=11
LCOORD=9
PPMEND=1.8
PPMST=4
IPAGE2 = 0
nameac(1) = 'Cre'
nameac(2) = 'Cho'
nameac(3) = 'NAA'
nameac(4) = 'NAAG'
nameac(5) = 'GPC'
nameac(6) = 'Glu'
nameac(7) = 'GSH'
nameac(8) = 'GABA'
nameac(9) = 'Gln'
nameac(10) = 'mI'
neach = 10
ISLICE = 1
IROWEN = 1
IRCOWST = 1

Concentration
relative to Cre and
CRLB

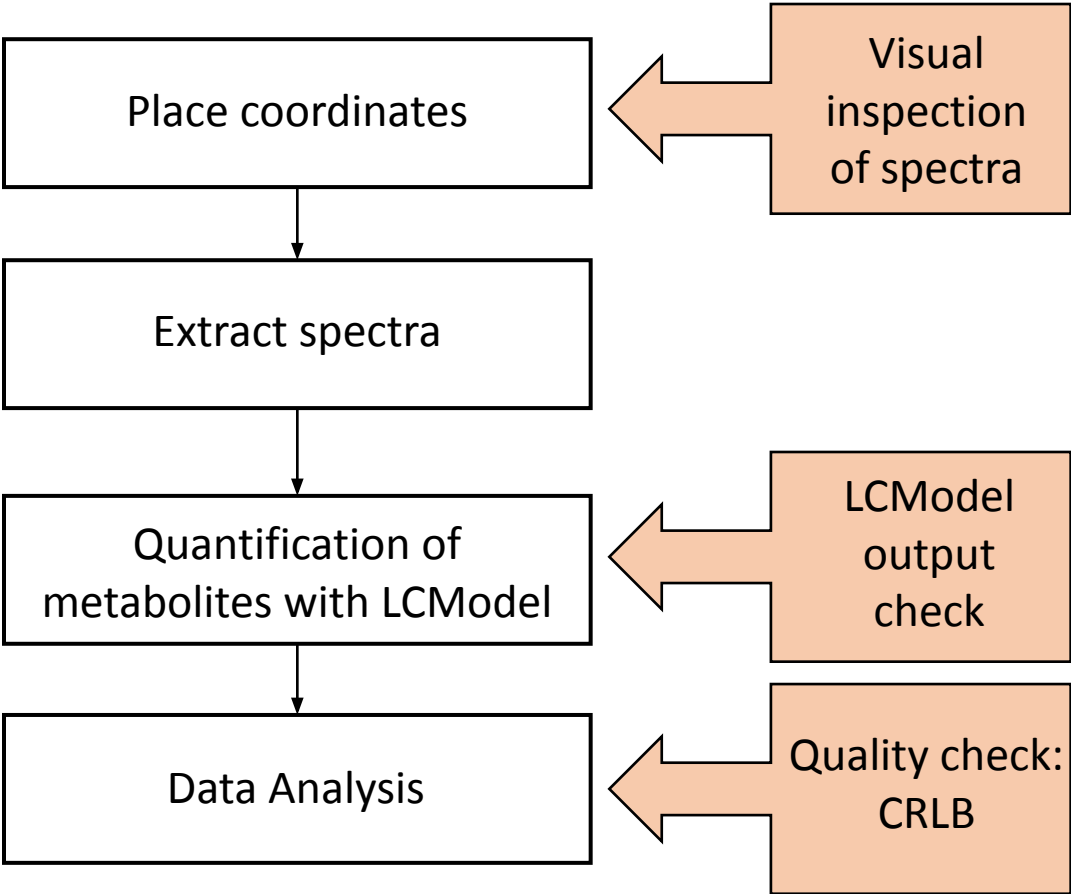
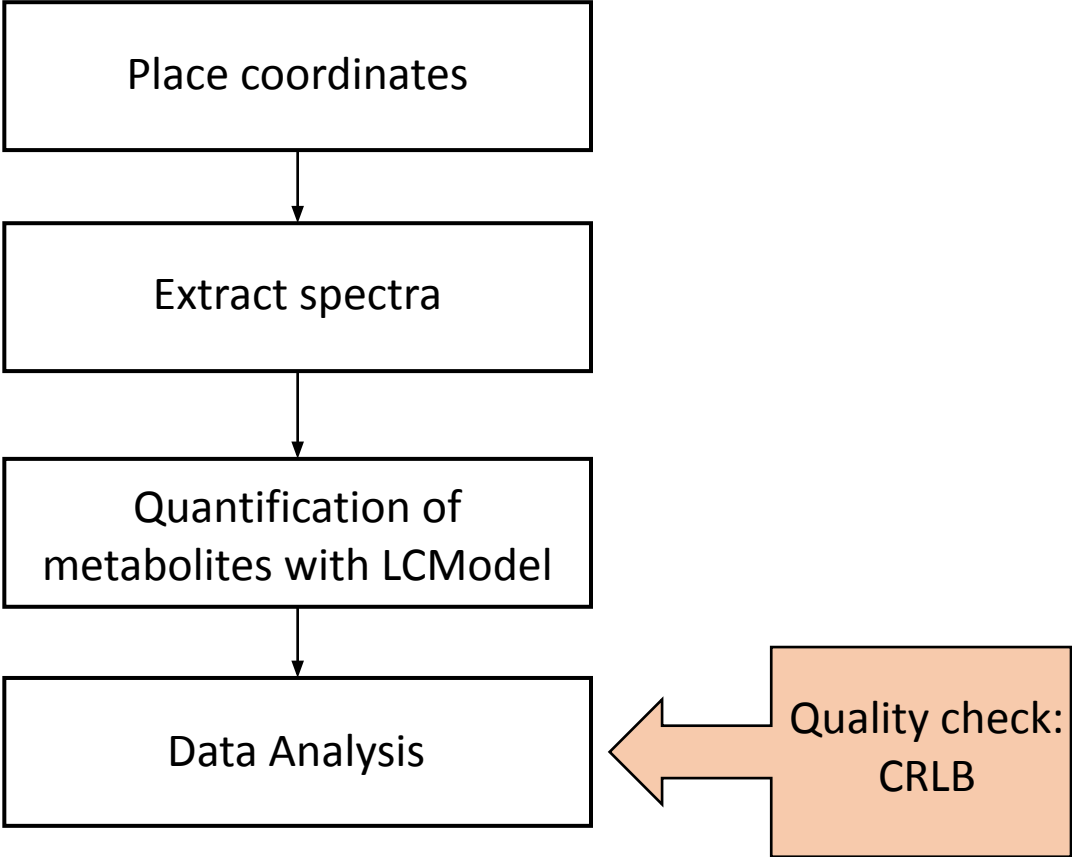
2. Model fit

3. Baseline

Current workflow



Updates



Proposed new quality check/ exclusionary criteria

Spectral inspection:

- Doubled peaks (motion)
- Lipid artifact
- Linewidth

LCModel check

- Model fit
- Baseline
- Residuals

Data

- $CRLB > 20$

Metabolites that we collect

- Aspartate
- Choline + GPC
- Cre
- GABA
- Glutamate
- Glutamine
- Glutathione
- Glucose

Disclaimer: we know very little about the actual biochemical roles of each of these (especially in humans), and even less about exactly what MRS picks up

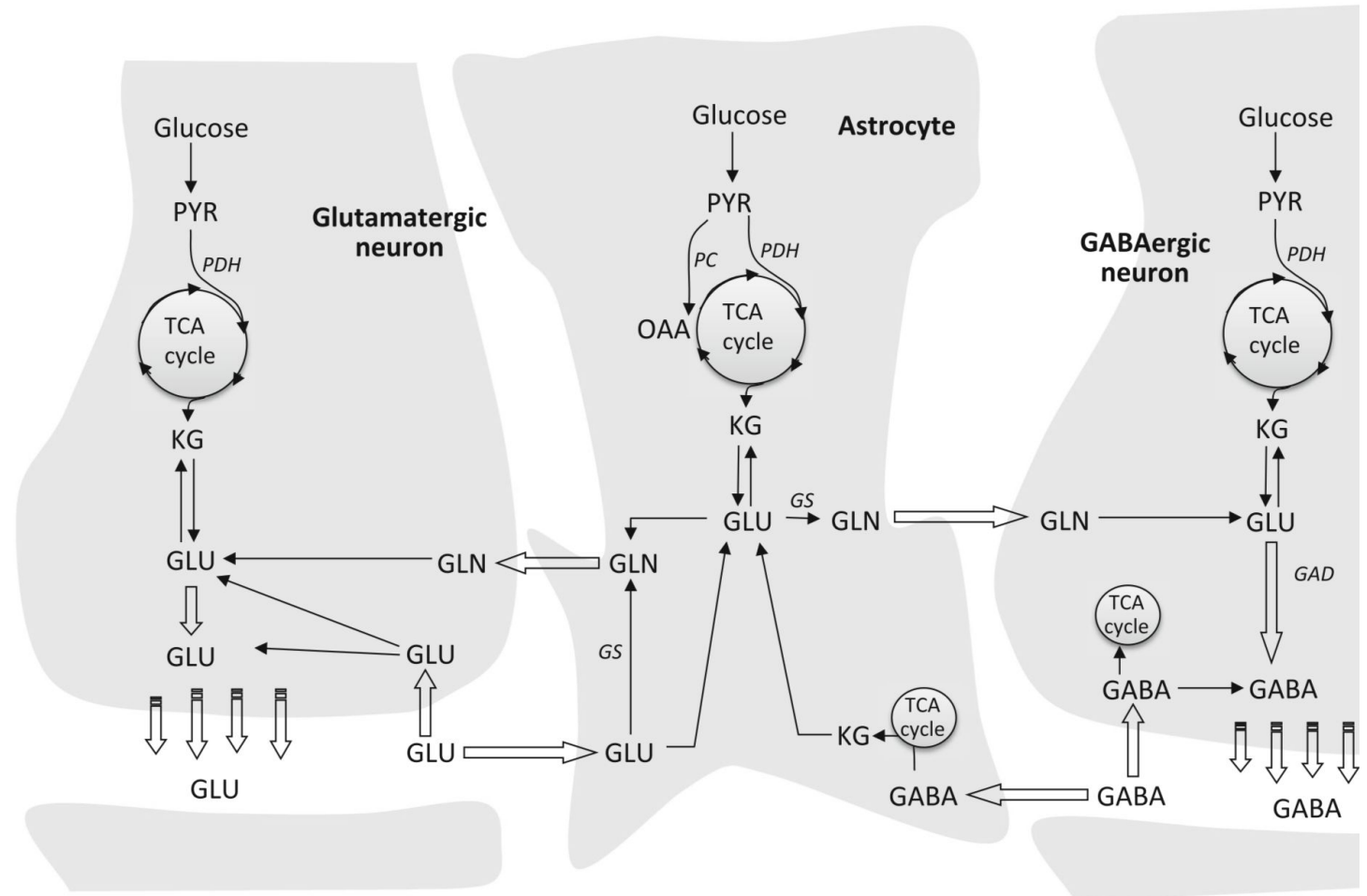
• NAA + NAAG

Creatine

- $Cr + PCr = tCr$
- Present in both neurons and glia
- Energy metabolism
- Use of Cr as ratio denominator
 - Correct for:
 - Stationarity assumption
 - Absolute quantification studies mixed/contradictory, but most find no changes
 - Introduces more variability than absolute metabolite quantification (Li et al., 2003)
 - Metabolite-to-creatine ratios estimated by LCModel more accurate than absolute (Kanowski et al., 2004)

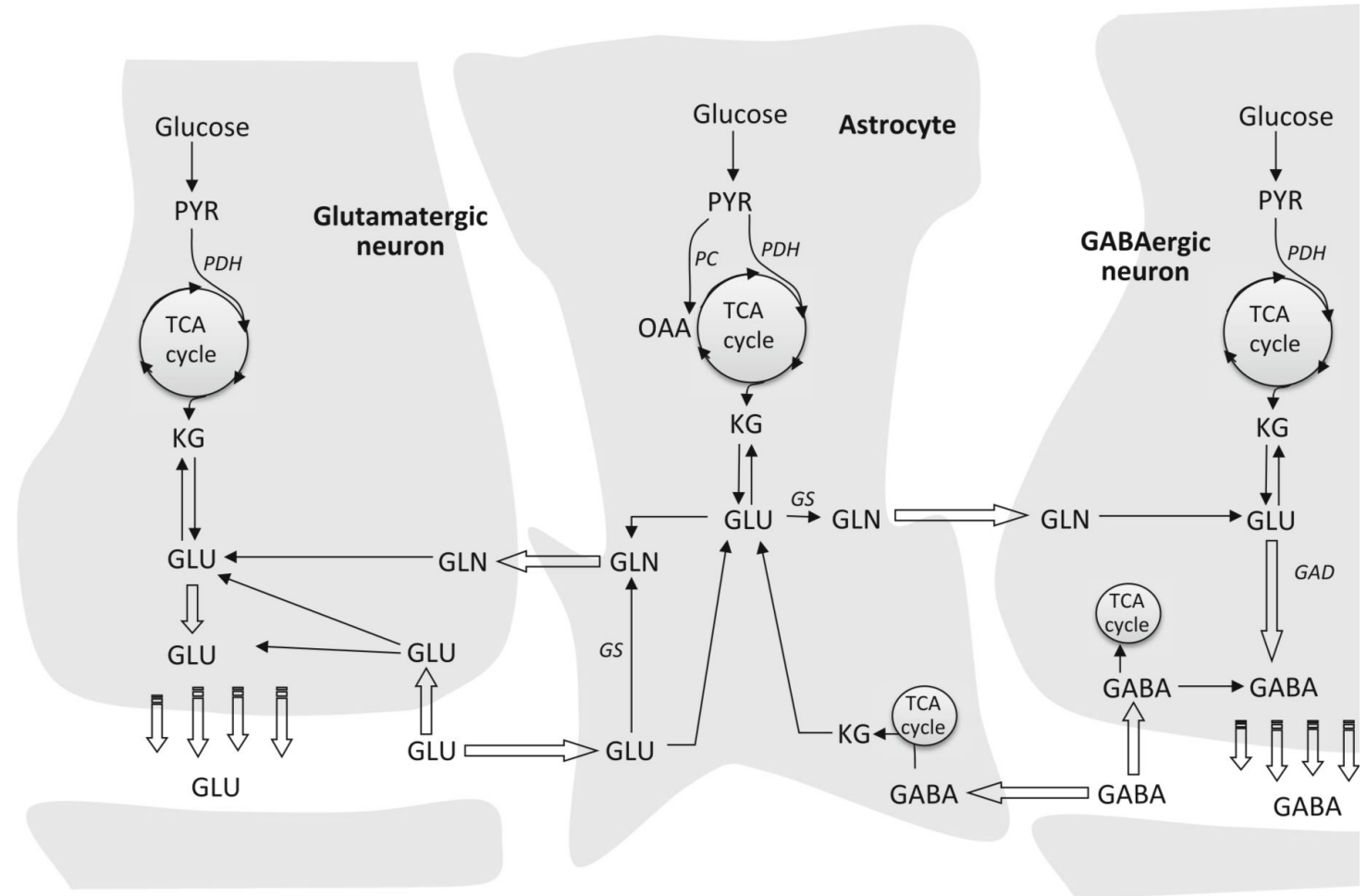
Glutamate and GABA

- In MRS: “Glutamate release and recycling is a major metabolic pathway that cannot be distinguished from its actions of neurotransmission.” (Rothman et al, 2002)
- Intracellular Glu: ~10mM; extracellular glu: 2 uM
- Intracellular GABA: 1 mM; extracellular GABA: 2 uM
- Exists in several metabolic pools
- Almost any pool may be available to be used as NT



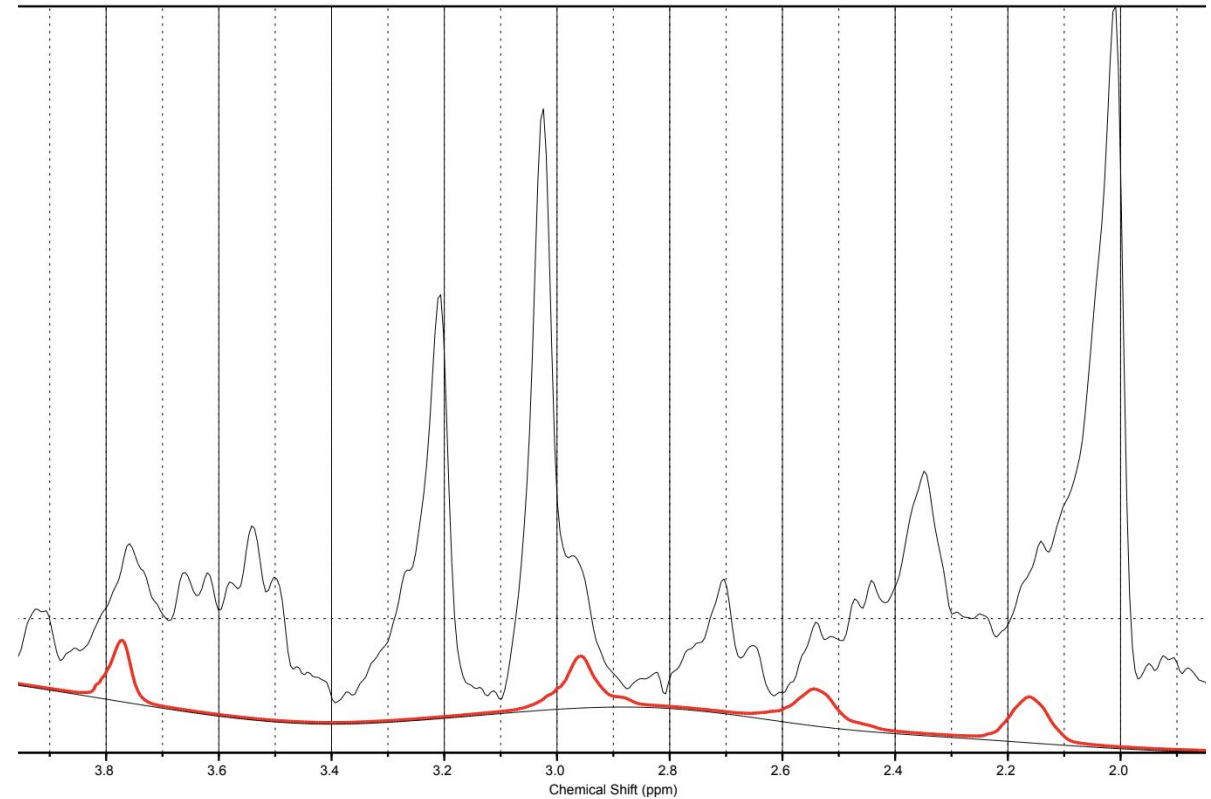
Glutamine

- Precursor to glutamate
- Glu stored as Gln in glia
- Small amounts of Gln also produced from GABA
- $\text{Glu} + \text{Gln} = \text{Glx}$



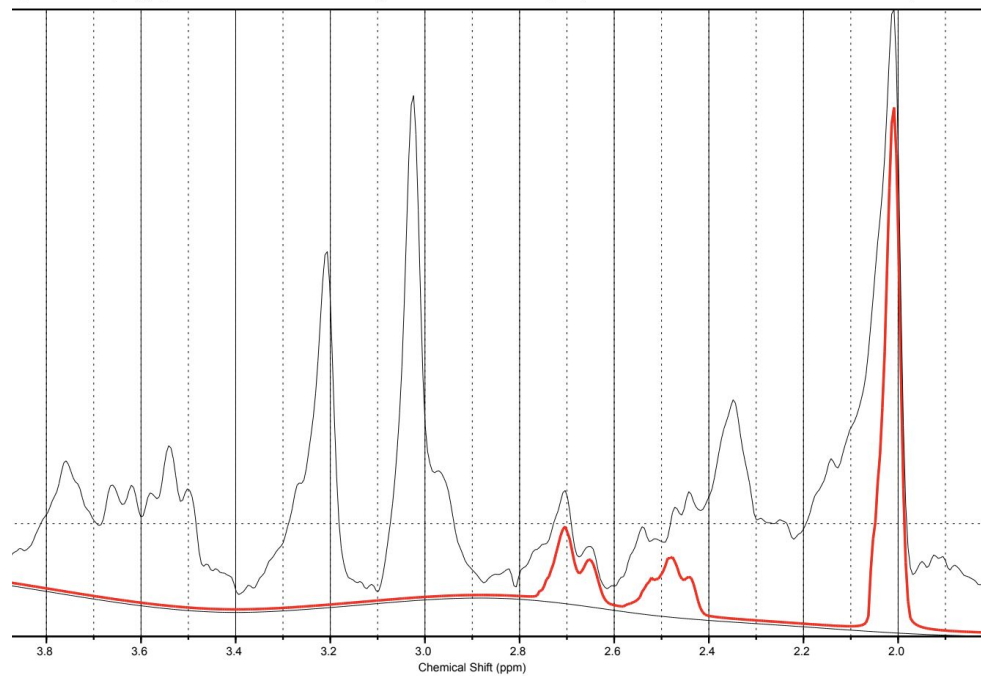
Glutathione

- Synthesized from glutamate
- Antioxidant, mitochondrial functions
- Generally elevated by stress
- Problematic to measure even at ultra-high field (14T)

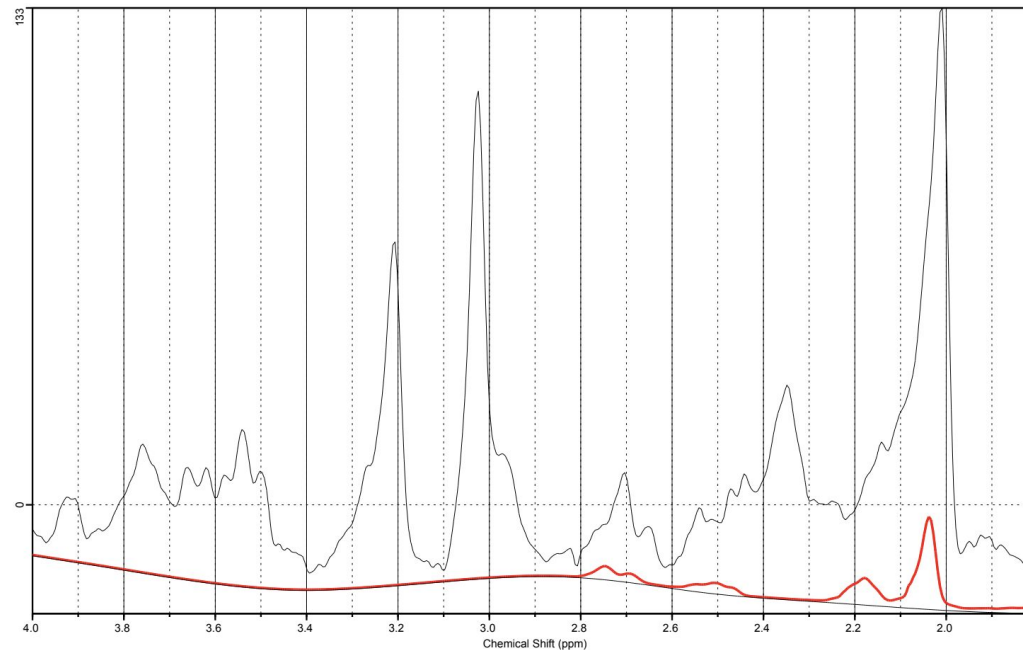


NAA + NAAG

- Role of NAA unclear; neuronal marker
 - Decreases with age
- NAAG can act as a transmitter; correlates with cognitive ability (Rahn et al., 2012)
- Separate measures likely unreliable (conversation with Victor, 2020)



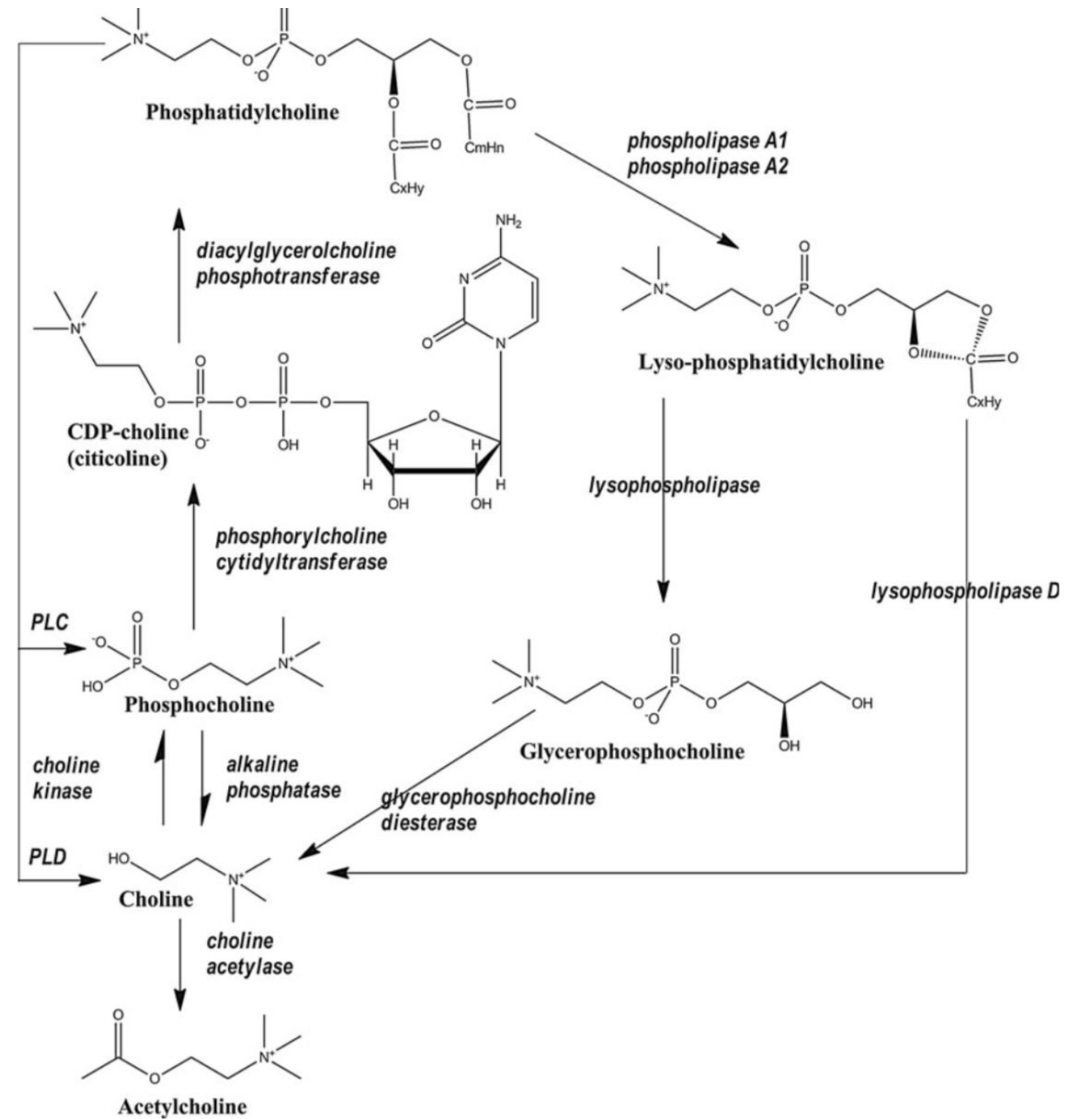
NAA



NAAG

Choline + GPC

- Role of Cho somewhat unclear
 - Membrane composition changes
- Cho correlated with acetylcholine
- Cho/PCho + GPC
 - Separate measures unreliable



Aspartate

- Amino acid
- Excitatory??
 - $^{-}\text{-(}\text{ツ}\text{)}\text{-}$
 - NMDA receptor agonist
 - Previously thought of as possibly excitatory NT, but recent evidence suggests otherwise (Herring et al., 2015)

Glucose

- Energy supply
- Levels of brain glucose related to levels of blood glucose more than to rate of brain glucose use (Rae, 2014)

Myoinositol

- Sugar synthesized from glucose
- Located mainly in glia
- Role:
 - membrane composition
 - second-messenger signaling pathways

Taurine

- Amino acid
- Role:
 - Osmoregulation
 - modulation of transmitter action via calcium